

#### **Thermal Lens Spectrometry: Theory and Applications**

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#### Outline

- 1. Basics of TLS spectrometry and comparison to UV-Vis spectrometry
- 2. Principles of collimated PB and multi-pass TLS instruments
- 3. Novel tunable TLS spectrometers
- 4. Limitations of multi-pass instruments
  - 1. Linearity range
  - 2. Effects of blank
- 5. Applications







#### **Basics of thermal lens effect**

- During non-radiative relaxation of excited species temperature in the sample increases (10<sup>-4</sup> 10<sup>-3</sup> K)
- a temperature gradient is generated with maximum temperature at the axis of the excitation beam
- the resulting refractive index gradient acts as a lens (mostly: dn/dT < 0, diverging lens)
- laser beam is defocused (single beam or pump/probe configuration)
- beam radius and its intensity at the beam axis changes
- relative change in the beam intensity is proportional to the absorbed power of the excitation beam.



#### TLS effect is time dependent - signal forms





• Absorbance (concentration), Power of excitation beam ( $I_0$ , usually denoted as P)  $I = I_0 10^{-A}$  $I = I_0 e^{-2.303A}$  $e^{A} = 1 + \frac{A}{11} + \frac{A^{2}}{21} + \frac{A^{3}}{21} + \dots$  $\frac{I}{I_0} = 1 - \frac{2.303A}{1!} + \frac{2.303^2A^2}{2!} - \frac{2.303^3A^3}{3!} + \dots$  $\frac{I-I_0}{I_0} = -\frac{2.303A}{1!} + \frac{2.303^2A^2}{2!} - \frac{2.303^3A^3}{3!} + \dots$ 

Absorbed power is linearly proportional to absorbance only for small A (< 0.1) !!!



- Optothermal parameters of sample/medium

   Thermal conductivity (k), temperature
   coefficient of refractive index (dn/dT)
- Probe beam wavelength ( $\lambda$ )  $\Theta = -\frac{2.303PA(\frac{dn}{dT})}{\lambda k}$
- Beam geometry factors: probe and pump beam radii ( $w_p$ ,  $w_e$ ), position of sample (z)

$$m = \left(\frac{w_p}{w_e}\right)^2, V = \frac{Z_1}{Z_c} + \frac{Z_c}{Z_2} \left[1 + \left(\frac{Z_1}{Z_c}\right)^2\right], \ Z_c = \frac{\pi w_{0p^2}}{\lambda}$$



#### **TLS experiment**





## Mathematical description of TLS effect/signal

TL causes a phase shift in propagation of probe beam

$$\begin{split} \Delta \Phi(r, z, t) &= -\frac{\Theta}{t_c(z)} \int_0^t \frac{dt'}{1 + 2t'/t_c(z)} \left[ 1 - \exp\left(-\frac{2r^2}{\omega_e^{-2}(z)(1 + 2t'/t_c(z))}\right) \right] \\ I(t) &= \\ &= I(0) \left\{ \left[ 1 - \frac{\Theta}{2} \arctan\left(\frac{2mV}{\left[(1 + 2m)^2 + V^2\right]\left(\frac{t_c}{t}\right) + 1 + 2m + V^2\right)}\right]^2 \right\} \\ &+ \left[ \frac{\Theta}{4} \ln\left(\frac{\left[1 + 2m/(1 + t/t_c)\right]^2 + V^2}{(1 + 2m)^2 + V^2}\right) \right]^2 \right\} \end{split}$$



#### Maximal TL signal

• *arctan* is maximum  $(\pi/2)$  when argument approaches  $\infty$ : highly collimated probe beam (no bimodal behavior)





#### Blank subtraction by differential TLS measurements





#### **E - Enhancement factor in TLS**

For spectrophotometry:  $\frac{I_0 - I}{I_0} = \frac{2.303A}{1!}$  therefore: for m = 1 and  $z_1 = z_c \sqrt{3}$ : and for max. signal:  $E = -\frac{P(\frac{dn}{dT})0.534}{\lambda k}$   $E_{coll} = -\frac{P(\frac{dn}{dT})\pi}{2\lambda k}$ 

Solvent	$-dn/dT / 10^{-4} \mathrm{K}^{-1}$	$k / W m^{-1} K^{-1}$	$E / 10^{-3} \mathrm{W}^{-1}$
H <sub>2</sub> O	0.91	0.607	0.12 (0.35)
CCl <sub>4</sub>	5.9	0.103	4.74 (13.9)
acetone	5.42	0.190	2.36 (6.94)

*E* is calculated for  $\lambda = 632.8$  nm (*E*<sub>coll</sub> is given in parentheses)

#### Adjustable beam size/position TLM



### **TLM detection in microfluidic systems**





### Efficiency of TLM for different sample thicknesses





#### **Multi-pass TLS instrument**





$$\Delta \Phi(r, z, t) = -n \frac{\Theta}{t_c(z)} \int_0^t \frac{dt'}{1 + 2t'/t_c(z)} \left[ 1 - \exp\left(-\frac{2r^2}{\omega_e^2(z)(1 + 2t'/t_c(z))}\right) \right]$$





### Milti-pass TLS measurements of Fe(II)-1,10 phenanthroline





#### **Blank effects**



This leads to 25-40% errors in 10-pass configuration for large  $\theta$ . e.g.  $\theta_b = 0.03$ for  $\theta_b = 0.01$  error is between 8 and 15% (A<sub>max</sub>= 0.05/cm) the effect increases with increasing concentration of analyte. It can be neglected for single and dual pass configurations.



#### TLS - advantages

- High sensitivity
  - signal proportional to excitation laser power
  - absorbances as low as 10<sup>-7</sup> can be measured
- Enables On-line detection
  - fast response of TLS signal (on µs to ms time scale)
- Capability of measuring small samples
  - sub-pL volumes can be probed
  - detection in microfluidic systems



#### TLS – drawbacks and solutions

- Sensitivity still needs improvement
  - Higher laser power? (photo-labile compounds)
  - Modify solvents
- Limited availability of laser sources
  - Coloring reactions, indirect detection
- Poor selectivity
  - Single wavelength measurements
  - Coupling to separation techniques (HPLC, IC, CE)
- Photodegradation
  - Measure in flowing systems



#### **Tunable Multi-pass TL**

#### spectrometer



 $P_e = 7 - 17 \text{ mW}$ , 10 nm FWHM, 4-pass configuration



#### **Recording TLS spectra with a multi-pass TL spectrometer**





550



#### **TLS spectra of luminescent**

#### nanogold materials





#### Determination of microcystin by PP2A inhibition assay

#### **Colorimetric reaction catalyzed by PP Reaction inhibited by cyanotoxin** Methyl-dehydro-alanine Iso-glutamic acid (Mdha) p-nitrophenol (Glu) phosphatase CH3 O COOH н OH OH Alanine H<sub>3</sub>C (colorless) OH CH2 (Ala) light н н OCH<sub>2</sub> vellow H H<sub>2</sub>C p-nitrophenilphosphate CH<sub>3</sub> HN H<sub>3</sub>C ĊH<sub>3</sub> CH3 Ĥ Ή Ô 0 3-amino-9-methoxy-2,6,8-trimethyl-COOH Leucine 10-phenyl-4.6-dienoic acid (Leu) p-nitrophenolate Methyl aspartic acid Objectiv Lens (MeAsp) ×/NA0.4 Sample > Waste Reagent To Objective Interference filter 200 Pinhole Mechanical Photodetector chopper He-Ne 95 10 **Argon ion laser** SR830 -45° 1.03 kH



#### **TLM-PP2A** inhibition assay



Enzyme consumption: 0.5 μL per injection Detection limit: ~ 80ng/L -12 times lower below the WHO limit for drinking water -8 times faster than batch mode assay





#### Transport phenomena in multiphase microflows - TLM Based Validation of Models



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flow in

### Detection and utilization of virus-like proteins and pseudovirions

HPV is a cause of cervical cancer. By using HPV antibodies one can detect HPV. By using HPV-VLPs one can detect HPV antibodies and past or current infection





#### Calibration curves for HPV-16 antibodies

Nanobeads

μFIA-TLM

(ng/mL)

 $6.8 \pm 0.9$ 

8 ± 1

 $5.4 \pm 0.2$ 

 $3.8 \pm 0.6$ 



#### **First detection and modulation of bilirubin in vascular endothelial cels**

![](_page_31_Figure_1.jpeg)

![](_page_32_Figure_0.jpeg)

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![](_page_33_Picture_12.jpeg)

![](_page_34_Picture_0.jpeg)

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![](_page_35_Picture_0.jpeg)

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